

Application News

High Performance Liquid Chromatograph Mass Spectrometer LCMS-9050

Rapid Screening Analysis of On-Line Purified Residual Pesticides in Crop Extract Using Accurate Mass Information

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User Benefits

- ◆ Enables analysis using positive/negative ion polarity switching by the LCMS-9050.
- Enables comprehensive analysis of compounds using the accurate mass and retention time information.
- ◆ This method using an online purification column enables quick and easy sample preparation.

■ Introduction

The pesticide residue limits in food are strictly regulated by each region or country in order to protect the people's health. In recent years, the number of pesticides regulated around the world is on the rise.

The QuEChERS method published by the USDA (United States Department of Agriculture) in 2003, which has been adopted as an official method by organizations such as the Association of Analytical Communities (AOAC) and Committee of European Normalization (CEN), enables efficient extraction of pesticides without the need for special equipment. However, there are problems with reproducibility being dependent on operators and the time required for the work.

In this article, an example of comprehensive analysis of residual pesticides in spinach using a Revive In-Line Sample Preparation (ILSP) column¹⁾ (RESTEK Co.) and the quadrupole time-of-flight mass spectrometer LCMS-9050 (Fig. 1) is introduced. The Revive ILSP column, which enables on-line cleanup of compounds derived from the matrix, makes the sample preparation quicker and easier. It is also expected that this method can contribute to the reduction of waste and costs associated with pretreatment.



Fig. 1 Exterior of LCMS-9050

■ Sample Preparation

Commercially available spinach and pesticides standard solution 74, 75 (Kanto Chemical Co., Inc.) were used for this analysis. The detailed preparation processes are shown in Fig. 2. 1.0 g of spinach that was frozen with dry ice and crushed was put in a 15 mL tube, and 3 mL of 1 % acetic acid in acetonitrile was added, then the tube was shaken. Subsequently, the tube was centrifuged after waiting for 2 minutes, and the supernatant was collected for sampling. Compared with the QuEChERS method, this method enables quicker and easier sample preparation. For example, it is expected that the time needed for sample preparation can be reduced to approximately one third by using this method when fourteen samples of spinach are pretreated¹⁾.

The recovery rate for losses in the preparation process and matrix effects were evaluated by adding a fixed concentration of pesticides standard solution to the spinach before extraction.

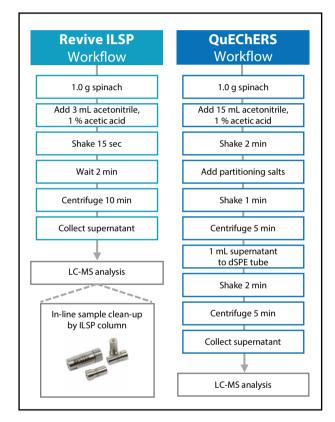


Fig. 2 Workflow for Sample Preparation (Left: Revive ILSP, Right: QuEChERS)

■ Analytical Conditions

For the analysis of pesticides, the method included in the LC/MS/MS Method Package Residual Pesticides Ver. 3 was applied to the LCMS-9050. The HPLC and MS conditions are shown in Table 1, and the outline of this system and the LC time program are shown in Figs. 3 and 4 respectively.

Hydrophobic compounds from the matrix in injected samples are separated from pesticides by the Revive ILSP column, and the pesticides are eluted to the analytical column (upper of Fig. 3). On the other hand, the compounds from the matrix are backflushed by a pump for washing and eluted to a drain, after switching a 6-port valve when the targeted pesticides have been eluted from the Revive ILSP column (bottom of Fig. 3 and Fig. 4). In this way, the Revive ILSP column can be reused.

Table 1 Analytical Conditions

UHPLC (Nexera [™] X3 syster	n)			
Analytical Column:	Shim-pack™ Velox Biphenyl			
	(100 mm L × 2.1 mm l.D., 2.7 μm) P/N: 227-32015-03			
In-Line Sample:	Revive ILSP Pesticides Single 5 \times 2.1 mm			
Preparation Column:	Cartridge (RESTEK)			
Mobile Phase A:	2 mM Ammonium formate-0.002 % Formic acid-Water			
Mobile Phase B, C (Wash):	2 mM Ammonium formate-0.002 % Formic acid-Methanol			
Gradient Program:	B conc. 3 % (0 min)-10 % (1 min)-55 % (3 min)- 100 % (10.5-12 min)-3 % (12.01-15 min)			
Flowrate (A & B):	0.4 mL/min			
Flowrate (C):	0 mL/min (0-5.49 min) – 1 mL/min (5.5-7.5 min) – 0.4 mL/min (7.51-11 min) - 0 mL/min (11.01-15			
Switching Valve Position:	min) load (0-6.49 min) – wash (6.5-10.5 min) – load			
Switching valve Fosition.	(10.51-15 min)			
Injection Volume:	2 μL (Co-injection 40 μL Water)			
MS (LCMS-9050)				
Ionization:	ESI (Positive, Negative)			
TOF-MS:	m/z 50-800			
Nebulizing Gas Flow:	2.0 L/min			
Drying Gas Flow:	10.0 L/min			
Heating Gas Flow:	10.0 L/min			
DL Temp.:	150 °C			
Block Heater Temp.:	300 °C			
Interface Temp.:	200 °C			
Probe Position:	+2 mm			

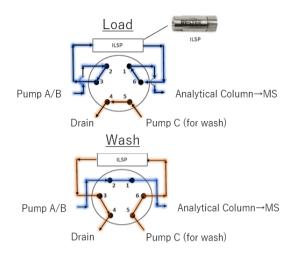


Fig. 3 Outline of ILSP System

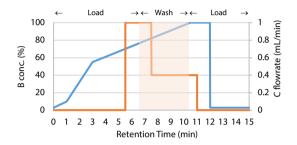


Fig. 4 LC Time Program

■ Analysis of Pesticides Standard Solution by the LCMS-9050

Table 2 shows the mass error of 79 compounds in pesticides standard solution diluted to 50 ppb. All the compounds were detected within \pm 1 mDa mass error. Theoretical m/z values of pesticides were calculated using LabSolutions Insight ExploreTM.

Fig. 5 shows the total ion current chromatogram (TICC) of the pesticides standard solution. Also, Fig. 6 shows the extracted ion chromatogram (XIC) of 79 compounds in pesticides standard solution and blank solvent.

Table 2 List of Pesticide Compounds

Compound	Molecular Formula	Selected	Theoretical		Retention Time	
·	C II Chi	lon	m/z	(mDa)	(min)	
Acetamiprid	C ₁₀ H ₁₁ CIN ₄	[M+H] ⁺	223.0745	-0.2	4.81	
Dimethoate	$C_5H_{12}NO_3PS_2$	$[M+H]^+$	230.0069	0.0	4.18	
Bromacil	$C_9H_{13}BrN_2O_2$	[M-H] ⁻	259.0087	0.4	4.80	
Propoxur	$C_{11}H_{15}NO_3$	$[M+H]^+$	210.1125	-0.2	5.01	
Isouron	$C_{10}H_{17}N_3O_2$	$[M+H]^+$	212.1394	-0.1	4.97	
Fluometuron	$C_{10}H_{11}F_3N_2O$	$[M+H]^+$	233.0896	-0.1	4.81	
Pyraclonil	$C_{15}H_{15}CIN_6$	[M+H] ⁺	315.1120	-0.1	7.07	
Metalaxyl/Metalaxyl-M*	$C_{15}H_{21}NO_4$	$[M+H]^+$	280.1544	-0.1	6.51	
Methidathion	$C_6H_{11}N_2O_4PS_3$	$[M+H]^+$	302.9692	-0.2	7.17	
Flumioxazin	$C_{19}H_{15}FN_2O_4$	$[M+H]^+$	355.1089	0.0	8.50	
Chlorbufam	$C_{11}H_{10}CINO_2$	$[M+H]^+$	224.0473	-0.5	5.91	
Ethiprole	$C_{13}H_9CI_2F_3N_4OS$	$[M+H]^+$	396.9899	0.3	5.95	
Paclobutrazol	$C_{15}H_{20}CIN_3O$	$[M+H]^+$	294.1368	-0.1	6.18	
Barban	$C_{11}H_9CI_2NO_2$	$[M+H]^+$	258.0083	-0.3	6.68	
Benthiavalicarb-isopropyl	$C_{18}H_{24}FN_3O_3S$	$[M+H]^+$	382.1595	-0.1	6.74	
Tiadinil	$C_{11}H_{10}CIN_3OS$	[M-H] ⁻	266.0160	0.0	6.55	
Triadimenol	$C_{14}H_{18}CIN_3O_2$	$[M+H]^+$	296.1161	-0.4	6.30	
Triflumizole Metabolite	$C_{12}H_{14}CIF_3N_2O$	$[M+H]^+$	295.0820	-0.2	4.81	
Prometryn	$C_{10}H_{19}N_5S$	$[M+H]^+$	242.1434	-0.2	6.66	
Tetraconazole	$C_{13}H_{11}CI_{2}F_{4}N_{3}O$	[M+H] ⁺	372.0288	-0.2	6.90	
Flusilazole	$C_{16}H_{15}F_2N_3Si$	[M+H] ⁺	316.1076	0.0	7.65	
Bensulide	$C_{14}H_{24}NO_4PS_3$	$[M+H]^+$	398.0678	-0.1	7.99	
Flubendiamide	$C_{23}H_{22}F_7IN_2O_4S$	[M-H] ⁻	681.0160	0.1	7.27	
Kresoxim-methyl	C ₁₈ H ₁₉ NO ₄	[M+H] ⁺	314.1387	-0.1	8.30	
Pyrazoxyfen	C ₂₀ H ₁₆ Cl ₂ N ₂ O ₃	[M+H] ⁺	403.0611	0.1	8.90	
Famoxadone	C ₂₂ H ₁₈ N ₂ O ₄	[M+NH ₄] ⁺	392.1605	0.1	8.60	
Phoxim	C ₁₂ H ₁₅ N ₂ O ₃ PS	[M+H] ⁺	299.0614	-0.2	8.47	
Trichlamide	C ₁₃ H ₁₆ Cl ₃ NO ₃	[M-H] ⁻	338.0123	0.6	7.40	
Metconazole	C ₁₇ H ₂₂ CIN ₃ O	[M+H] ⁺	320.1524	-0.1	7.56	
Pyraclofos	C ₁₄ H ₁₈ CIN ₂ O ₃ PS	[M+H] ⁺	361.0537	0.1	8.44	

Compound	Molecular Formula	Selected Ion	Theoretical m/z	Mass Error (mDa)	Retention Time (min)
Bitertanol	C ₂₀ H ₂₃ N ₃ O ₂	[M+H] ⁺	338.1863	-0.2	7.94
Pyrazophos	$C_{14}H_{20}N_3O_5PS$	$[M+H]^+$	374.0934	0.0	9.38
Diflufenican	$C_{19}H_{11}F_5N_2O_2$	[M+H] ⁺	395.0814	0.0	8.26
Pentoxazone	C ₁₇ H ₁₇ CIFNO ₄	$[M+NH_4]^+$	371.1168	0.1	9.24
Tolfenpyrad	$C_{21}H_{22}CIN_3O_2$	[M+H] ⁺	384.1473	0.0	9.08
Pyributicarb	$C_{18}H_{22}N_2O_2S$	$[M+H]^+$	331.1475	0.1	9.47
Chlorpyrifos	$C_9H_{11}CI_3NO_3PS$	$[M+H]^{+}$	349.9336	-0.2	9.25
Etoxazole	$C_{21}H_{23}F_2NO_2$	[M+H] ⁺	360.1770	0.1	9.39
Cyenopyrafen	$C_{24}H_{31}N_3O_2$	[M+H] ⁺	394.2489	0.1	9.45
Spirodiclofen	$C_{21}H_{24}CI_2O_4$	[M+H] ⁺	411.1125	-0.1	9.62
3-Hydroxycarbofuran	C ₁₂ H ₁₅ NO ₄	[M+H] ⁺	238.1074	-0.1	4.06
Cymoxanil	$C_7H_{10}N_4O_3$	[M+H] ⁺	199.0826	-0.4	4.35
Phosphamidon	$C_{10}H_{19}CINO_5P$	[M+H] ⁺	300.0762	-0.1	5.04
Terbacil	$C_9H_{13}CIN_2O_2$	[M-H] ⁻	215.0593	-0.2	4.86
XMC (3,5-xylyl methylcarbamate)	$C_{10}H_{13}NO_2$	[M+H] ⁺	180.1019	-0.2	5.25
Flutriafol	$C_{16}H_{13}F_2N_3O$	[M+H] ⁺	302.1100	0.1	
Fensulfothion	C ₁₁ H ₁₇ O ₄ PS ₂	[M+H] ⁺	309.0379	0.0	
Triforine (isomer-1)	C ₁₀ H ₁₄ Cl ₆ N ₄ O ₂	[M+H] ⁺	432.9321	0.2	
Triforine (isomer-2)	C ₁₀ H ₁₄ Cl ₆ N ₄ O ₂	[M+H] ⁺	432.9321	0.4	
Diethofencarb	C ₁₄ H ₂₁ NO ₄	[M+H] ⁺	268.1544	-0.3	
Fludioxonil	C ₁₂ H ₆ F ₂ N ₂ O ₂	[M-H] ⁻	247.0324	0.2	
Mandipropamid	$C_{23}H_{22}CINO_4$	[M+H] ⁺	412.1310	0.0	
Pyriminobac-methyl (E)	$C_{17}H_{19}N_3O_6$	[M+H] ⁺	362.1347	-0.1	
Malathion	C ₁₀ H ₁₉ O ₆ PS ₂	[M+H] ⁺	331.0434	-0.1	
Bromobutide-dibromo	C ₁₅ H ₂₂ BrNO	[M+H] ⁺	234.1852	0.0	
Fluopicolide	C ₁₄ H ₈ Cl ₃ F ₃ N ₂ O	[M+H] ⁺	382.9727	0.1	
Triadimefon	C ₁₄ H ₁₆ CIN ₃ O ₂	[M+H] ⁺	294.1004	-0.1	
Flamprop-methyl	C ₁₇ H ₁₅ CIFNO ₃	[M+H] ⁺	336.0797	-0.1	
Bromobutide	C ₁₅ H ₂₂ BrNO	[M+H] ⁺	312.0958	0.0	
Carfentrazone-ethyl	C ₁₅ H ₁₄ Cl ₂ F ₃ N ₃ O ₃	[M+NH ₄] ⁺	429.0702	0.1	
Dimethametryn	C ₁₁ H ₂₁ N ₅ S	[M+H] ⁺	256.1591	-0.2	
Penthiopyrad	C ₁₆ H ₂₀ F ₃ N ₃ OS	[M+H] ⁺	360.1352	0.0	
Tebuconazole	C ₁₆ H ₂₂ CIN ₃ O	[M+H] ⁺	308.1524	0.0	
Benalaxyl	C ₂₀ H ₂₃ NO ₃	[M+H] ⁺	326.1751	0.0	
Oxadiargyl	C ₁₅ H ₁₄ Cl ₂ N ₂ O ₃	[M+H] ⁺	341.0454	-0.2	
Isoxathion	C ₁₃ H ₁₆ NO ₄ PS	[M+H] ⁺	314.0611	0.0	
Prochloraz	C ₁₅ H ₁₆ Cl ₃ N ₃ O ₂	[M+H] ⁺	376.0381	0.0	
Pirimiphos-methyl	C ₁₁ H ₂₀ N ₃ O ₃ PS	[M+H] ⁺	306.1036	-0.1	
Difenoconazole	C ₁₉ H ₁₇ Cl ₂ N ₃ O ₃	[M+H] ⁺	406.0720	-0.5	
Trifloxystrobin	C ₂₀ H ₁₉ F ₃ N ₂ O ₄	[M+H] ⁺	400.0720	0.3	
Triflumizole	C ₂₀ 11 ₁₉ 1311 ₂ O ₄ C ₁₅ H ₁₅ ClF ₃ N ₃ O	[M+H] ⁺	346.0929	0.0	
Amisulbrom	$C_{13}H_{13}BrFN_5O_4S_2$	[M+H] ⁺	467.9628	0.3	
Profenofos	C ₁₃ H ₁₅ BrClO ₃ PS	[M+H] ⁺	374.9402	-0.2	
Buprofezin		[M+H] ⁺			
•	C ₁₆ H ₂₃ N ₃ OS		306.1635	0.0	
Piperonyl butoxide	C ₁₉ H ₃₀ O ₅	[M+NH ₄] ⁺	356.2432	-0.2	
Butachlor	C ₁₇ H ₂₆ CINO ₂	[M+H] ⁺	312.1725	-0.1	
Quinoxyfen	C ₁₅ H ₈ Cl ₂ FNO	[M+H] ⁺	308.0040	0.0	
Pyridaben	C ₁₉ H ₂₅ CIN ₂ OS	[M+H] ⁺	365.1449	0.2	
Fenpropimorph	C ₂₀ H ₃₃ NO	[M+H] ⁺	304.2635	-0.2	7.10

*Pesticides standard solution contains 50 ppb each of metalaxyl and metalaxyl-M.

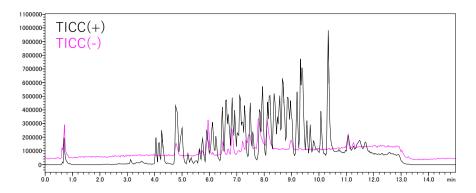


Fig. 5 Total Ion Current Chromatogram (TICC) of Pesticides Standard Solution

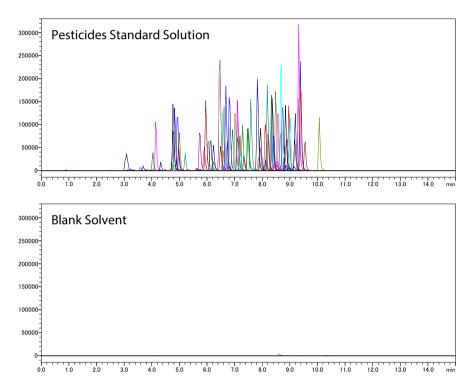
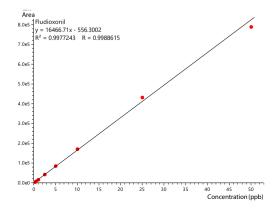


Fig. 6 Extracted Ion Chromatograms (XICs) of 79 Pesticide Compounds

■ Linearity of Calibration Curve

Linearity of the calibration curve for each pesticide was evaluated by generating a 7-point calibration curve with the range 0.5-50 ppb in solvent (acetonitrile) and in spinach extract. Both in solvent and in extract, linearity showed very good results (coefficient of determination R²: 0.99 or more) for all the

compounds. For 68 out of 79 compounds detection was within 2.5 ppb or less. Calibration curves for fludioxonil, which was detected in negative mode, in solvent and in extract are shown in Fig. 7 as an example, and calibration ranges for all the compounds are shown in Table 3.



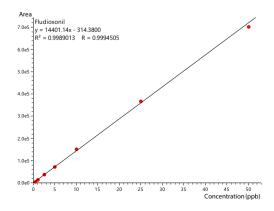


Fig. 7 Calibration Curve of Fludioxonil (Left: in Solvent, Right: in Spinach Extract)

Table 3 Linear Ranges of 79 Pesticides

Compound		on Range pb)		Calibration Range (ppb)	
	in solvent	in spinach extract	Compound	in solvent	in spinach extract
Acetamiprid	1-50	2.5-50	3-Hydroxycarbofuran	1-50	1-50
Dimethoate	1-50	0.5-50	Cymoxanil	5-50	5-50
Bromacil	5-50	5-50	Phosphamidon	0.5-50	0.5-50
Propoxur	1-50	1-50	Terbacil	5-50	5-50
Isouron	0.5-50	0.5-50	XMC (3,5-xylyl methylcarbamate)	2.5-50	2.5-50
Fluometuron	0.5-50	0.5-50	Flutriafol	1-50	1-50
Pyraclonil	0.5-50	0.5-50	Fensulfothion	0.5-50	0.5-50
Metalaxyl/Metalaxyl-M*	0.5-50	0.5-50	Triforine (isomer-1)	10-50	10-50
Methidathion	1-50	1-50	Triforine (isomer-2)	10-50	10-50
Flumioxazin	5-50	5-50	Diethofencarb	1-50	1-50
Chlorbufam	25-50	25-50	Fludioxonil	0.5-50	0.5-50
Ethiprole	1-50	1-50	Mandipropamid	0.5-50	0.5-50
Paclobutrazol	1-50	1-50	Pyriminobac-methyl (E)	0.5-50	0.5-50
Barban	5-50	5-50	Malathion	0.5-50	0.5-50
Benthiavalicarb-isopropyl	0.5-50	0.5-50	Bromobutide-debromo	1-50	1-50
Tiadinil	1-50	1-50	Fluopicolide	0.5-50	1-50
Triadimenol	2.5-50	2.5-50	Triadimefon	1-50	1-50
Triflumizole Metabolite	0.5-50	0.5-50	Flamprop-methyl	0.5-50	0.5-50
Prometryn	0.5-50	0.5-50	Bromobutide	1-50	2.5-50
Tetraconazole	0.5-50	0.5-50	Carfentrazone-ethyl	1-50	1-50
Flusilazole	0.5-50	0.5-50	Dimethametryn	0.5-50	0.5-50
Bensulide	1-50	1-50	Penthiopyrad	0.5-50	0.5-50
Flubendiamide	1-50	2.5-50	Tebuconazole	1-50	0.5-50
Kresoxim-methyl	0.5-50	1-50	Benalaxyl	0.5-50	0.5-50
Pyrazoxyfen	0.5-50	0.5-50	Oxadiargyl	2.5-50	2.5-50
Famoxadone	2.5-50	2.5-50	Isoxathion	0.5-50	0.5-50
Phoxim	1-50	0.5-50	Prochloraz	1-50	1-50
Trichlamide	2.5-50	2.5-50	Pirimiphos-methyl	0.5-50	0.5-50
Metconazole	1-50	0.5-50	Difenoconazole	1-50	0.5-50
Pyraclofos	0.5-50	0.5-50	Trifloxystrobin	0.5-50	0.5-50
Bitertanol	2.5-50	2.5-50	Triflumizole	0.5-50	0.5-50
Pyrazophos	0.5-50	0.5-50	Amisulbrom	25-50	25-50
Diflufenican	1-50	1-50	Profenofos	0.5-50	0.5-50
Pentoxazone	10-50	10-50	Buprofezin	0.5-50	0.5-50
Tolfenpyrad	0.5-50	1-50	Piperonyl butoxide	0.5-50	0.5-50
Pyributicarb	0.5-50	0.5-50	Butachlor	5-50	5-50
Chlorpyrifos	1-50	1-50	Quinoxyfen	0.5-50	1-50
Etoxazole	0.5-50	0.5-50	Pyridaben	0.5-50	0.5-50
Cyenopyrafen	0.5-50	0.5-50	Fenpropimorph	0.5-50	0.5-50
Spirodiclofen	1-50	1-50			

*Metalaxyl and metalaxyl-M were not distinguished.

■ Spike and Recovery Test

A spike and recovery test was performed using spinach extract to which 79 pesticides standard solution was spiked at 0.01 mg/kg per sample (concentration in pretreated sample solution was 2.5 ppb). The recovery rate was calculated at the concentration of 2.5 ppb, if not possible, at the concentration of 50 ppb. The results of recovery rate calculated by external standard method and reproducibility (n=5) are shown in Table 4, and the breakdown of the recovery rate is shown in Fig. 8.

Recovery rates were 70-120 % for 77 of the 79 compounds and %RSDs were less than 20 % for all the compounds. Good recovery rate and reproducibility were obtained without significant matrix inhibition by using the Revive ILSP column in sample preparation.

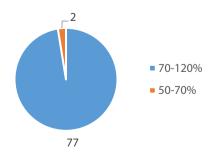


Fig. 8 Breakdown of Recovery Rate

Table 4 Recovery Rate and Reproducibility (%RSD) (n=5)

Compound Recovery %RSD Compound		Compound	Recovery Rate (%)	%RSD	
Acetamiprid	82.6	11.4	3-Hydroxycarbofuran	93.4	14.6
Dimethoate	87.1	7.6	Cymoxanil	81.9	2.6
Bromacil	73.1	2.5	Phosphamidon	96.8	6.5
Propoxur	103.9	9.9	Terbacil	74.9	3.7
Isouron	107.1	1.5	XMC (3,5-xylyl methylcarbamate)	88.2	7.7
Fluometuron	105.4	4.7	Flutriafol	99.6	8.4
Pyraclonil	107.8	5.6	Fensulfothion	104.6	4.0
Metalaxyl/Metalaxyl-M*	108.6	3.6	Triforine (isomer-1)	97.9	7.5
Methidathion	110.5	10.3	Triforine (isomer-2) Diethofencarb	101.8 100.2	4.3 6.8
Flumioxazin	65.5	1.7	Fludioxonil	88.4	4.5
Chlorbufam	85.0	12.7	Mandipropamid	104.5	3.2
Ethiprole	104.3	7.9	Pyriminobac-methyl (E)	100.3	3.5
Paclobutrazol	103.2	8.4	Malathion	105.2	5.1
Barban	92.1	4.0	Bromobutide-debromo	107.0	12.2
Benthiavalicarb-isopropyl	108.2	1.3	Fluopicolide	96.2	7.2
Tiadinil	99.3	7.1	Triadimefon	96.5	6.0
Triadimenol	100.4	18.6	Flamprop-methyl	105.6	8.4
Triflumizole Metabolite	103.9	4.0	Bromobutide	112.5	9.9
Prometryn	107.2	5.7	Carfentrazone-ethyl	77.3	6.0
Tetraconazole	98.1	5.1	Dimethametryn	106.5	5.2
Flusilazole	97.2	5.9	Penthiopyrad	97.9	5.7
Bensulide	102.3	3.6	Tebuconazole	109.9	6.8
Flubendiamide	95.3	14.4	Benalaxyl	93.3	3.0
Kresoxim-methyl	95.9	9.9	•	66.0	12.0
Pyrazoxyfen	116.9	4.9	Oxadiargyl Isoxathion	94.3	3.9
Famoxadone	84.6	18.6			
Phoxim	85.2	12.5	Prochloraz	98.3	6.2
Trichlamide	80.7	17.2	Pirimiphos-methyl	100.4	5.1
Metconazole	95.2	5.3	Difenoconazole	95.7	7.7
Pyraclofos	115.7	5.9	Trifloxystrobin	96.7	3.1
Bitertanol	77.4	6.4	Triflumizole	83.4	3.1
Pyrazophos	112.3	2.1	Amisulbrom	98.1	4.4
Diflufenican	83.6	5.6	Profenofos	92.1	3.2
Pentoxazone	97.9	3.1	Buprofezin	94.1	4.1
Tolfenpyrad	101.3	4.6	Piperonyl butoxide	87.6	7.6
Pyributicarb	102.9	2.6	Butachlor	100.9	3.0
Chlorpyrifos	104.2	13.3	Quinoxyfen	100.4	3.2
Etoxazole	108.0	2.1	Pyridaben	95.0	4.9
Cyenopyrafen	98.8	4.7	Fenpropimorph	103.7	3.9
Spirodiclofen	110.5	6.7			

*Metalaxyl and metalaxyl-M were not distinguished.

■ Conclusion

Screening analysis of pesticides was performed using positive/negative ion polarity switching by the LCMS-9050. The on-line sample preparation method with a Revive ILSP column made it possible to speed up and simplify the preparation process. Screening analysis of pretreated spinach samples with pesticides standard spiked provided good results for recovery rate, reproducibility and linearity. For sensitivity, 68 out of 79 compounds were detected at 2.5 ppb or less. This method is applicable for the analysis of residual pesticides in other crops.

<References>

1) Lupo, S.A., Romesberg, R.L., Lu, X., 2020. Automated inline pigment removal for the analysis of pesticide residues in spinach by liquid chromatography tandem mass spectrometry, J. Chromatogr. A 1629, 461477.

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