

Application News

No. B88

Probe Electrospray Ionization Mass Spectrometer

Establishment of a Method for Direct Analysis of the Mouse Liver Metabolome Using the DPiMS™-8060

In the analysis of endogenous metabolites (metabolome analysis), it is difficult to perfectly remove biases caused by pretreatment and sampling. Therefore, in order to accurately grasp the changes in the metabolome of a biospecimen, the establishment of a method for direct analysis of the metabolome is indispensable. Probe electrospray ionization (PESI) is a new direct ionization method in which an ultrafine and minimally invasive probe is used for sampling. Acquired samples are ionized by applying a high voltage to the probe tip and therefore components can be analyzed without using a chromatograph.

By using the DPiMS-8060 probe electrospray ionization tandem mass spectrometer (Fig. 1), which combines PESI with tandem mass spectrometry, direct analysis of the metabolome of a biospecimen is possible.

This article introduces a method established to directly analyze the metabolome of a tissue sample (intact metabolome analysis) using a PESI tandem mass spectrometer together with the application of the method to the metabolome analysis of CCl₄-induced acute liver failure model mice.

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Fig. 1 DPiMS™-8060

■ Sample Preparation and Analytical Conditions

Standard metabolite samples including amino acids, organic acids, and sugars (26 metabolites) were prepared by diluting them with 50 % ethanol solution and dripping 10 μL of each sample into dedicated sample plates for liquid samples (Shimadzu Corporation). We then selected the MRM transitions for each compound and optimized the mass spectrometer conditions such as collision energy (CE). The information on the optimized MRM transitions of the 26 metabolites is listed in Table 1.

Next, liver samples were collected from mice by dissection. Square sections about 3 mm in size were taken from a healthy mouse and liver failure model mouse (liver failure induced by administering carbon tetrachloride) each. The sections were placed in dedicated sample plates for solid samples and then set on the instrument. Since solid samples can be analyzed by the DPiMS-8060 by simply placing them in a sample plate, there is no need for complex pretreatment.

Table 1 MRM Transitions of 26 Metabolites

Name	Polarity	Transition (m/z)	Collision Energy (V)
3-hydroxybutyrate	(-)	103.1>59.0	35
Citric acid/isocitric acid	(-)	191.0>111.1	20
D-glucose	(-)	179.1>59.2	20
Glucose-6-phosphate	(-)	259.1>96.9	20
Glutaric acid	(-)	131.0>87.3	20
Glycine	(-)	74.2>74.2	20
L-asparagine	(-)	131.0>113.3	20
L-aspartic acid	(-)	131.9>88.1	20
L-glutamic acid	(-)	146.0>102.1	20
L-lactic acid	(-)	89.0>43.2	20
L-malic acid	(-)	133.0>114.9	20
L-serine	(-)	103.9>74.2	20
Pyruvic acid	(-)	87.1>43.1	20
Succinic acid	(-)	117.1>73.0	20
Taurine	(-)	124.0>80.0	20
2-aminobutyric acid	(+)	104.1>58.1	20
L-glutamine	(+)	147.1>84.2	20
L-histidine	(+)	156.1>110.3	20
L-leucine/L-isoleucine	(+)	132.1>86.2	20
L-methionine	(+)	150.3>104.1	20
L-ornithine	(+)	132.9>70.0	20
L-phenylalanine	(+)	166.2>120.2	20
L-proline	(+)	116.2>70.0	20
L-threonine	(+)	120.1>74.0	20
L-tryptophan	(+)	205.2>146.1	20
L-tyrosine	(+)	182.1>136.1	20

